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## Ferulates, amurenlactone A and amurenamide A from traditional Chinese medicine cortex *Phellodendri Amurensis*

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Two new ferulate compounds, amurenlactone A (**1**) and amurenamide A (**2**), along with 11 known compounds have been isolated from the cortex of *Phellodendron amurense*. The structures of the new compounds were established based on 1D, 2D NMR and mass spectral analyzes. The known compounds were identified by comparison with authentic samples.

**Keywords:** *Phellodendron amurense*; Rutaceae; cortex; amurenlactone A; amurenamide A

### 1. Introduction

The traditional Chinese medicine, cortex *Phellodendri Amurensis*, has been used for the treatment of bacillary dysentery, inflammation, tuberculosis and liver cirrhosis.<sup>1,2</sup> Several reports have been encountered on the isolation of isoquinoline alkaloids, phenolic compounds, flavonoids, limonoidal triterpenes and sterols from this plant.<sup>2–6</sup> However, most of the plant materials of these researches were obtained from the species growing in Japan, instead of China. Therefore, the cortex of *Phellodendron amurense* obtained from Liaoning province, a famous region yielding cortex *Phellodendri Amurensis* in China, was reinvestigated and resulted in the isolation of two new ferulate compounds amurenlactone A (**1**) and amurenamide A (**2**), along with 11 known compounds. We report herein the isolation and structural elucidation of the new compounds using chromatographic and spectral methods.

### 2. Results and discussion

Amurenlactone A (**1**) was obtained as white amorphous powder,  $[\alpha]_D^{25} - 6.8$ . Both  $\text{FeCl}_3$  and ferric hydroxamate tests showed positive results which indicated that compound **1** has a phenolic hydroxyl and a lactone group. The molecular formula was determined as  $\text{C}_{17}\text{H}_{20}\text{O}_9$  by HR-ESI-MS at  $m/z$  369.1184  $[\text{M} + \text{H}]^+$ . The IR absorption bands at 3420 and  $1786\text{ cm}^{-1}$  were consistent with the presence of a hydroxyl group and a  $\gamma$ -lactone ring. The  $^1\text{H}$  NMR spectrum displayed one set of *trans*-double bond signals at  $\delta$  7.69 (1H, d,  $J = 16.0\text{ Hz}$ , H-7') and 6.41 (1H, d,  $J = 16.0\text{ Hz}$ , H-8'). One set of ABX signals at  $\delta$  7.20 (1H, d,  $J = 1.5\text{ Hz}$ , H-2'), 7.11 (1H, dd,  $J = 8.0, 1.5\text{ Hz}$ , H-6') and 6.81 (1H, d,  $J = 8.0\text{ Hz}$ , H-5') were attributed to a 1,3,4-trisubstituted benzene ring. The HMQC correlations between H-7' and C-7' and the HMBC correlations from H-2' to C-6', C-7' indicated that the *trans*-double bond was connected to the C-1' of the benzene ring

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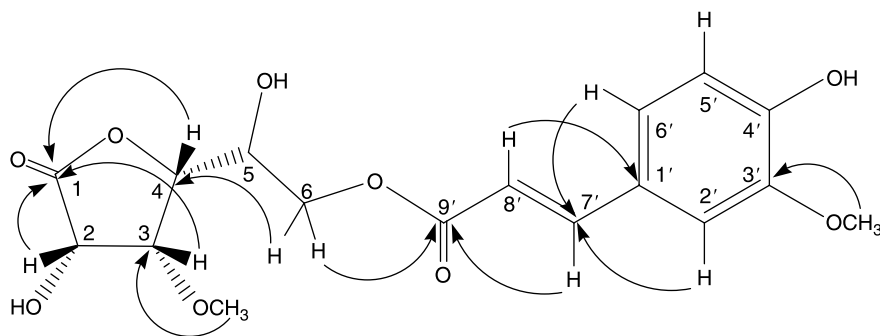


Figure 1. The selected HMBC correlations for compound **1**.

(Figure 1). The HMBC correlations between the methoxy proton at  $\delta$  3.89 (3H, *s*) and C-3', and H-7' and the carbonyl carbon C-9' at  $\delta$  169.0, suggested that **1** has a feruloyl moiety. There were four oxymethine proton signals at  $\delta$  4.65 (1H, *d*,  $J = 4.4$  Hz, H-2), 4.11 (1H, *dd*,  $J = 4.4, 3.3$  Hz, H-3), 4.47 (1H, *dd*,  $J = 8.0, 3.3$  Hz, H-4) and 4.19 (1H, *m*, H-5), and one oxymethylene proton signal at  $\delta$  4.30 (2H, *m*, H-6) in the  $^1\text{H}$  NMR spectrum. The  $^1\text{H}$ - $^1\text{H}$  COSY spectrum showed that these protons correlated in turn, and their corresponding carbons were confirmed by HMQC spectrum (Table 1). The carbonyl carbon signal at  $\delta$  177.4 (C-1) in  $^{13}\text{C}$  NMR spectrum and the HMBC correlations between C-1 and H-2, H-3, and H-4 (Figure 1), combined with the IR absorption band at  $1786\text{ cm}^{-1}$ , indicated that compound **1** should include a hexano- $\gamma$ -lactone moiety. The methoxy at  $\delta$  3.56 (3H, *s*) was connected to C-3 according to the HMBC correlation between the methoxy and C-3. The feruloyl moiety was linked to C-6 since the long-range correlation of H-6 with C-9' was displayed in the HMBC spectrum (Figure 1). Therefore, **1** was considered to have the structure of 6-*O*-feruloyl-hexano- $\gamma$ -lactone.

Pei-Lin Wu *et al.* isolated a compound vittarilide-A from *Vittaria anguste-elongata* with similar structure as **1** except that the substituting groups of C-3 and 3' in vittarilide-A are hydroxyls,<sup>7</sup> and its  $^{13}\text{C}$  NMR spectral data were similar to those of **1** except for C-3 (Table 1). In vittarilide-A, the H-3 *trans* to H-2 and *cis* to H-4 were proven by the larger

coupling constant of  $J_{\text{H-2/H-3}} = 9.3$  Hz and the smaller coupling constant of  $J_{\text{H-3/H-4}} = 5.0$  Hz. Comparing with vittarilide-A, the smaller coupling constant of  $J_{\text{H-2/H-3}} = 4.4$  Hz and  $J_{\text{H-3/H-4}} = 3.3$  Hz were observed in **1**, which suggested that the relative configuration of H-3 to H-2 and H-4 is all *cis*. Correlations between H-2 and H-3, H-3 and H-4 in NOESY experiment also proved that these three protons projected in the same direction. The similar coupling constants of  $J_{\text{H-2/H-3}}$ ,  $J_{\text{H-3/H-4}}$  and  $J_{\text{H-4/H-5}}$  compared with mannono- $\gamma$ -lactone, as well as the negative optical rotation of **1**, indicated that it is a derivative of L-mannono- $\gamma$ -lactone.<sup>8,9</sup> Therefore, **1** was determined as 6-*O*-feruloyl-3-*O*-methyl-L-mannono- $\gamma$ -lactone, named as amurenolactone A (**1**).

Amurenamide A (**2**) was obtained as yellow oil,  $[\alpha]_{\text{D}}^{25} - 4.4$ .  $\text{FeCl}_3$  test showed positive results which indicated that compound **2** has phenolic hydroxyl group. The molecular formula was determined as  $\text{C}_{17}\text{H}_{23}\text{O}_9\text{N}$  by HR-ESI-MS at  $m/z$  386.1446  $[\text{M} + \text{H}]^+$ . The IR absorption band at  $1674$  ( $\nu_{\text{C=O}}$ ),  $1631$  ( $\nu_{\text{NH}}$ ) and  $1429$  ( $\nu_{\text{C-N}}$ )  $\text{cm}^{-1}$  suggested the presence of primary amide (because of the existence of hydroxyl group in **2**, the absorption band of  $\nu_{\text{NH}}$  was not shown in the IR spectrum). The low field of the  $^1\text{H}$  NMR spectrum was similar to compound **1**, which displayed that **2** has the same feruloyl fragment as **1**. The  $^1\text{H}$ - $^1\text{H}$  COSY spectrum showed correlations of four oxymethine protons at  $\delta$  4.44 (1H, *d*,

Table 1. <sup>1</sup>H and <sup>13</sup>C NMR spectral data (δ) of compounds 1–2 (CD<sub>3</sub>OD), and vittarilide-A (acetone-d<sub>6</sub>).

| No.                 | 1              |                                       |                         | 2              |   |                         | vittarilide-A  |      |                |
|---------------------|----------------|---------------------------------------|-------------------------|----------------|---|-------------------------|----------------|------|----------------|
|                     | δ <sub>C</sub> | δ <sub>H</sub>                        | HMBC                    | δ <sub>C</sub> | δ <sub>H</sub>  | HMBC                    | δ <sub>C</sub> | HMBC | δ <sub>C</sub> |
| 1                   | 177.4          |                                       |                         | 175.8          |   |                         | 175.3          |      |                |
| 2                   | 72.9           | 4.65 (1H, d, <i>J</i> = 4.4 Hz)       | C-1                     | 72.9           | 4.44 (1H, d, <i>J</i> = 2.5 Hz)   | C-1                     | 74.5           |      |                |
| 3                   | 80.0           | 4.11 (1H, dd, <i>J</i> = 3.3, 4.4 Hz) | C-1, 3-OCH <sub>3</sub> | 83.4           | 3.74 (1H, t, <i>J</i> = 3.1 Hz)   | C-1, 3-OCH <sub>3</sub> | 74.5           |      |                |
| 4                   | 81.5           | 4.47 (1H, dd, <i>J</i> = 3.3, 8.0 Hz) | C-1, C-3, C-5           | 72.3           | 3.90 (1H, t, <i>J</i> = 3.1 Hz)   |                         | 80.1           |      |                |
| 5                   | 69.4           | 4.19 (1H, m)                          |                         | 70.7           | 4.03 (1H, m)  | C-3                     | 69.0           |      |                |
| 6                   | 65.4           | 4.30 (2H, m)                          | C-4, C-9'               | 66.3           | 4.19 (1H, dd, <i>J</i> = 6.6, 11.1 Hz) 4.28 (1H, dd, <i>J</i> = 4.3, 11.1 Hz) | C-4, C-9'               | 66.4           |      |                |
| 1'                  | 127.7          |                                       |                         | 127.5          |   |                         | 127.5          |      |                |
| 2'                  | 111.9          | 7.20 (1H, d, <i>J</i> = 1.5 Hz)       | C-3', C-4', C-6', C-7'  | 111.4          | 7.34 (1H, d, <i>J</i> = 1.5 Hz)   | C-3', C-4', C-6', C-7'  | 115.1          |      |                |
| 3'                  | 149.4          |                                       |                         | 148.7          |   |                         | 146.2          |      |                |
| 4'                  | 150.7          |                                       |                         | 150.0          |   |                         | 148.7          |      |                |
| 5'                  | 116.5          | 6.81 (1H, d, <i>J</i> = 8.0 Hz)       | C-1', C-3', C-4', C-6'  | 116.0          | 6.87 (1H, d, <i>J</i> = 8.0 Hz)   | C-1', C-3', C-4', C-6'  | 116.3          |      |                |
| 6'                  | 124.2          | 7.11 (1H, dd, <i>J</i> = 1.5, 8.0 Hz) | C-2', C-4', C-7'        | 123.9          | 7.14 (1H, dd, <i>J</i> = 1.5, 8.0 Hz)   | C-2', C-4', C-7'        | 122.5          |      |                |
| 7'                  | 147.4          | 7.69 (1H, d, <i>J</i> = 16.0 Hz)      | C-1', C-9'              | 145.8          | 7.61 (1H, d, <i>J</i> = 16.0 Hz)  | C-1', C-9'              | 146.0          |      |                |
| 8'                  | 115.1          | 6.41 (1H, d, <i>J</i> = 16.0 Hz)      | C-1', C-9'              | 115.8          | 6.38 (1H, d, <i>J</i> = 16.0 Hz)  | C-1', C-9'              | 115.3          |      |                |
| 9'                  | 169.0          |                                       |                         | 167.5          |   |                         | 167.5          |      |                |
| 3-OCH <sub>3</sub>  | 60.8           | 3.56 (3H, s)                          | C-3                     | 59.0           | 3.48 (3H, s)  | C-3                     |                |      |                |
| 3'-OCH <sub>3</sub> | 56.5           | 3.89 (3H, s)                          | C-3'                    | 56.3           | 3.90 (3H, s)  | C-3'                    |                |      |                |

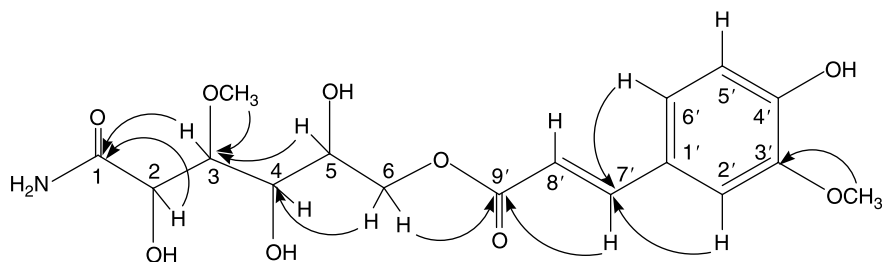


Figure 2. The selected HMBC correlations for compound **2**.

$J = 2.5$  Hz, H-2), 3.74 (1H, t,  $J = 3.1$  Hz, H-3), 3.90 (1H, t,  $J = 3.1$  Hz, H-4) and 4.03 (1H, m, H-5) successively, and the correlations of the oxymethylene protons at  $\delta$  4.19 (1H, dd,  $J = 6.6, 11.1$  Hz, H-6 $\alpha$ ) and 4.28 (1H, dd,  $J = 4.3, 11.1$  Hz, H-6 $\beta$ ) with H-5 were also observed. Only H-2 and H-3 correlated with the carbonyl carbon at  $\delta$  175.8 (C-1) in the HMBC spectrum indicating that this structural fragment in **2** should be a straight chain, which was also supported by the unsaturation degree of **2**. The methoxy protons at  $\delta$  3.48 (3H, s) correlated with C-3 in HMBC spectrum, so it should be connected to C-3. Since the correlation of H-6 with C-9' was observed in the HMBC spectrum, the feruloyl should be connected to C-6 (Figure 2). According to the molecular formula, it is concluded that there is an  $-\text{NH}_2$  group in **2**, and IR spectrum showed the absorption for a primary amide group; therefore, **2** was determined as 6-*O*-feruloyl-3-methoxy-2,4,5-trihydroxycaproylamine, named as amurenamide A. The relative configurations of C-2, C-3, C-4 and C-5 were not determined.

In addition, 11 compounds obakunone (**3**), obaculactone (**4**), kihadanin B (**5**), berberine (**6**),  $\beta$ -sitosterol (**7**), daucosterol (**8**), palmatine (**9**), syringin (**10**), methyl 3-*O*-feruloyl quinate (**11**), jatrorrhizine (**12**) and sanleng acid (**13**) were also isolated from the cortex of *P. amurense*.

### 3. Experimental

#### 3.1 General experimental procedures

UV spectra were measured on a Hitachi UV-3210 spectrophotometer; IR spectra were

measured on a Bruker Is-55 spectrophotometer as KBr disks. Optical activities were measured on a Perkin-Elmer 241MC polarimeter. NMR spectra were recorded on Bruker ARX-300 spectrometers, using tetramethylsilane as internal standard; all chemical shifts are reported in parts per million (ppm,  $\delta$ ). ESI mass spectra were recorded on a LCQ LC-MS spectrometer and HR-ESI-MS spectra were recorded on a ZAB-HS GC-MS spectrometer. Column chromatography was carried out on silica gel (100–140 and 200–300 mesh; Qingdao Haiyang Chemical Co., Shandong, China) and Sephadex LH-20 (Sigma, USA). Pre-coated TLC plates were used without activation (silica gel 60 F254, 0.25 mm, Merck, Germany).

#### 3.2 Plant material

The cortex of *P. amurense* were collected in September 1999 from Tieling, Liaoning province, China, and authenticated by Professor Zheng Cui of Shenyang Pharmaceutical University, China. A voucher specimen (No. SPU-PA-99-9) has been deposited in the School of Traditional China Materia Medica, Shenyang Pharmaceutical University, Shenyang, China.

#### 3.3 Extraction and isolation

The air-dried and powdered cortex of *P. amurense* (7.2 kg) was extracted three times with 70% ethanol under reflux. The extract was concentrated under reduced pressure to give dark brown syrup. The syrup was successively partitioned with petroleum

ether, chloroform, and *n*-butanol to obtain chloroform extract 120 g and *n*-butanol extract 276 g. Compounds **3–7** and **13** were isolated from chloroform extract by repeated chromatography on silica gel column.

The *n*-butanol extract was chromatographed over silica gel column using a gradient of chloroform–methanol (100:1 to 100% methanol) to afford six fractions (PBFr I–VI). PBFr II was subjected to column chromatography over silica gel with chloroform–methanol (10:1) followed by Sephadex LH-20 chromatography with methanol, and further purified by preparative TLC with chloroform–methanol–ammonia (10:1:0.5) to yield **2** (10 mg). PBFr III was chromatographed over silica gel using a gradient of chloroform–methanol to afford three fractions (PBFr III-1, -2 and -3). PBFr III-1 was subjected to column chromatography over silica gel with cyclohexane and ethyl acetate (1:1) and rechromatographed over silica gel column with chloroform–methanol (20:1), yielding **1** (7.7 mg). Compounds **8–11** and **12** were also isolated from *n*-butanol extract by column chromatography on silica gel or Sephadex LH-20 and preparative TLC successively.

### 3.3.1 Amurenlactone A (**1**)

C<sub>17</sub>H<sub>20</sub>O<sub>9</sub>, white amorphous powder.  $[\alpha]_D^{25} - 6.8$  (*c* 0.0051, MeOH). UV (MeOH)  $\lambda_{\max}$  nm (log  $\epsilon$ ): 204 (3.97), 233 (3.79), 326 (3.94). IR  $\nu_{\max}$  cm<sup>-1</sup> 3420 (OH), 1786 (C=O). <sup>1</sup>H NMR and <sup>13</sup>C NMR spectral data (Table 1). Negative ESI-MS (probe)

12 eV, *m/z* (rel. int.): 367 [M – H]<sup>-</sup> (76), 335 [M – H<sub>2</sub>O–Me]<sup>-</sup> (100), 233 (14). HR-ESI-MS *m/z* 369.1184 [M + H]<sup>+</sup> (calcd for C<sub>17</sub>H<sub>21</sub>O<sub>9</sub>, 369.1180).

### 3.3.2 Amurenamide A (**2**)

C<sub>17</sub>H<sub>23</sub>O<sub>9</sub>N, yellow oil,  $[\alpha]_D^{25} - 4.4$  (*c* 0.0037, MeOH). UV (MeOH)  $\lambda_{\max}$  nm (log  $\epsilon$ ): 202 (3.70), 218 (3.63), 324 (3.62). IR  $\nu_{\max}$  cm<sup>-1</sup> 3422 (OH), 1674 (NH<sub>2</sub>–C=O). <sup>1</sup>H NMR and <sup>13</sup>C NMR spectral data (Table 1). Positive ESI-MS (probe) 12 eV, *m/z* (rel. int.): 386 [M + H]<sup>+</sup>(50), 368 [M – NH<sub>3</sub>]<sup>+</sup>(100). HR-ESI-MS *m/z* 386.1446 [M + H]<sup>+</sup> (calcd for C<sub>17</sub>H<sub>24</sub>O<sub>9</sub>N, 386.1446).

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